

Comprehensive Cancer Center
Tübingen-Stuttgart

Post ASH 2024 San Diego



EBERHARD KARLS
UNIVERSITÄT
TÜBINGEN



 Comprehensive
Cancer Center
Tübingen - Stuttgart

 **Universitätsklinikum**
Tübingen

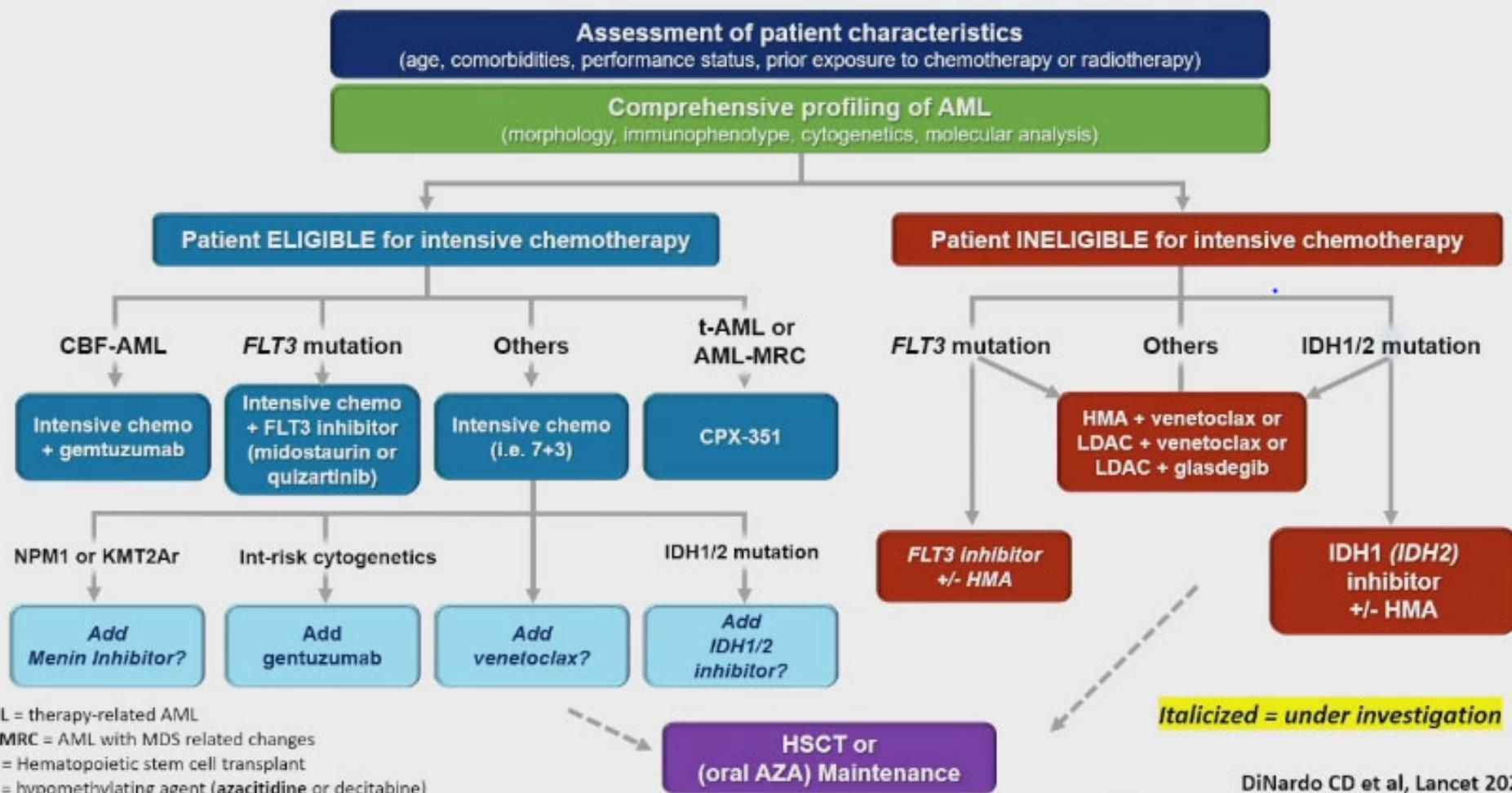
Update on diagnostics – Post ASH 2024

1. Education Program AML M&Ms: How to Integrate Mutations and MRD Data
2. Artificial Intelligence in Hematology: From generative AI to Ethics and Applications
3. #1538 Optical Genome Mapping As Standard-of-Care in Acute Leukemia: Diagnostic and Clinical Impacts 10 Months Post-Implementation

Latest Classifications for Acute Myeloid Leukemia and Myelodysplastic Syndromes

1. WHO 5th Edition Khoury JD, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Myeloid and Histiocytic/Dendritic Neoplasms. *Leukemia*.
2. ICC 2022 Arber DA et al. International Consensus Classification of Myeloid Neoplasms and Acute Leukemias: integrating morphologic, clinical, and genomic data. *Blood*. 2022
3. ELN 2022 Hartmut Döhner, et al. Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood* 2022

Evolving diagnostic and treatment paradigm for Newly Dx AML



DiNardo CD et al, Lancet 2024

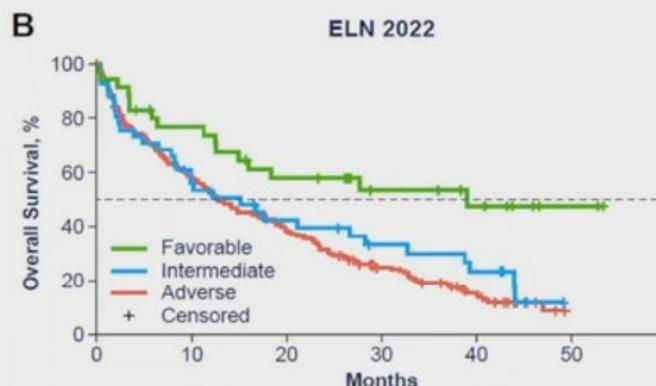


Neither ELN 2017 nor 2022 discriminate optimally for patients receiving HMA+VEN therapies



Patients at Risk						
46	28	20	12	10	2	
65	44	29	17	9	0	
168	90	58	31	14	0	

ELN 2017	n	Events	Median OS, months
			(95% CI)
Favorable	46	25	21.1 (9.9, NR)
Intermediate	65	48	23.3 (12.9, 28.3)
Adverse	168	141	11.5 (8.9, 16.2)



Patients at Risk						
35	25	18	11	8	2	
41	22	15	10	7	0	
203	115	74	39	18	0	

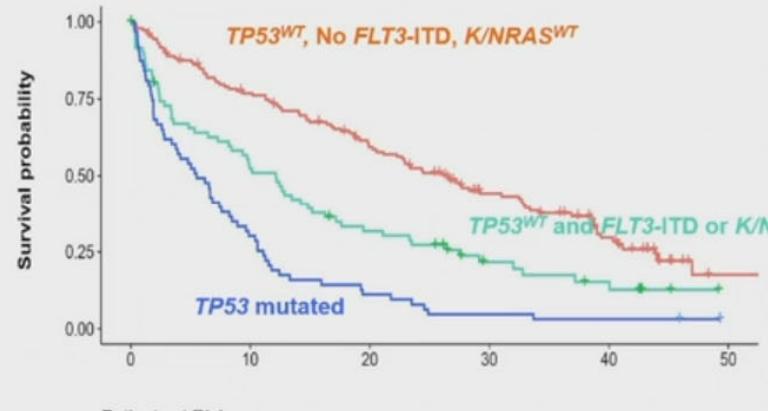
ELN 2022	n	Events	Median OS, months
			(95% CI)
Favorable	35	16	39.0 (12.5, NR)
Intermediate	41	31	15.2 (7.9, 28.3)
Adverse	203	167	12.7 (10.4, 17.6)

Dohner H et al, Blood 2024



Patients receiving Ven+Aza are **better characterized** by three molecularly-defined subgroups

"mPRS Score"

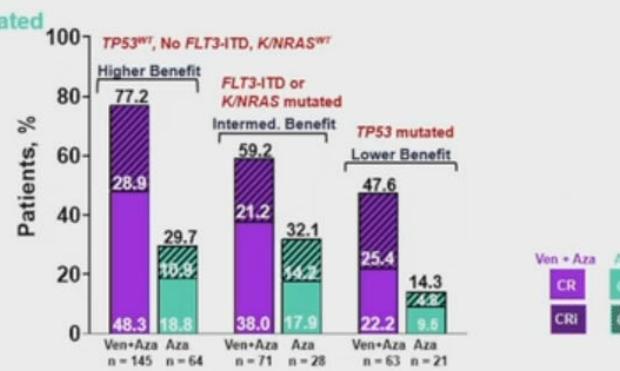


Benefit Group

	Higher Benefit	Interm. Benefit	Lower Benefit
Patients at Risk	145	107	79
	107	36	21
	47	10	10
	25	6	3
	2	0	2

mPRS = modified prognostic risk signature

Ven + Aza (N = 279)	n	Events	Median OS, months (95% CI)
Higher Benefit	145	96	26.51 (20.24, 32.69)
Intermediate Benefit	71	57	12.12 (7.26 – 15.15)
Lower Benefit	63	61	5.52 (2.79 – 7.59)



Dohner H et al, Blood 2024



Latest Classifications for Acute Myeloid Leukemia and Myelodysplastic Syndromes

1. WHO 5th Edition Khoury JD, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Myeloid and Histiocytic/Dendritic Neoplasms. *Leukemia* 2022
2. ICC 2022 Arber DA et al. International Consensus Classification of Myeloid Neoplasms and Acute Leukemias: integrating morphologic, clinical, and genomic data. *Blood* 2022
3. ELN 2022 Hartmut Döhner, et al. Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood* 2022
4. ELN 2024 LIT Hartmut Döhner, et al. Döhner H, DiNardo CD, Appelbaum FR, et al. Genetic risk classification for adults with AML receiving less-intensive therapies: the 2024 ELN recommendations. *Blood* 2024

Risk Classifications Updated to Reflect Intended Therapy

ELN 2022, Döhner et al, *Blood*, 2022

Favorable	<ul style="list-style-type: none"> t(8;21)(q22;q22.1)/RUNX1::RUNX1T1 inv(16)(p13.1q22) or t(16;16)(p13.1;q22)/CBFB::MYH11 Mutated NPM1 without <i>FLT3</i>-ITD bZIP in-frame mutated <i>CEBPA</i>
Intermediate	<ul style="list-style-type: none"> Mutated <i>NPM1</i> with <i>FLT3</i>-ITD Wild-type <i>NPM1</i> with <i>FLT3</i>-ITD (without adverse-risk genetic lesions) t(9;11)(p21.3;q23.3)/MLLT3::KMT2A Cytogenetic and/or molecular abnormalities not classified as favorable or adverse
Adverse	<ul style="list-style-type: none"> t(6;9)(p23.3;q34.1)/DEK::NUP214 t(v;11q23.3)/KMT2A-rearranged t(9;22)(q34.1;q11.2)/BCR::ABL1 t(8;16)(p11.2;p13.3)/KAT6A::CREBBP inv(3)(q21.3q26.2) or t(3;3)(q21.3;q26.2)/<i>GATA2</i>, <i>MECOM</i>(<i>EVI1</i>) t(3q26.2;v)/<i>MECOM</i>(<i>EVI1</i>)-rearranged -5 or del(5q); -7; -17/abn(17p) Complex karyotype, ** monosomal karyotype†† Mutated <i>ASXL1</i>, <i>BCOR</i>, <i>EZH2</i>, <i>RUNX1</i>, <i>SF3B1</i>, <i>SRSF2</i>, <i>STAG2</i>, <i>U2AF1</i>, and/or <i>ZRSR2</i> Mutated <i>TP53</i>

ELN 2024 LIT, Döhner et al, *Blood*, 2024

Favorable	<ul style="list-style-type: none"> Mutated <i>NPM1</i> (<i>FLT3</i>-ITD^{neg}, <i>NRAS</i>^{wt}, <i>KRAS</i>^{wt}, <i>TP53</i>^{wt}) Mutated <i>IDH2</i> (<i>FLT3</i>-ITD^{neg}, <i>NRAS</i>^{wt}, <i>KRAS</i>^{wt}, <i>TP53</i>^{wt}) Mutated <i>IDH1</i>* (<i>TP53</i>^{wt}) Mutated <i>DDX41</i>†, Other cytogenetic and/or molecular abnormalities‡ (<i>FLT3</i>-ITD^{neg}, <i>NRAS</i>^{wt}, <i>KRAS</i>^{wt}, <i>TP53</i>^{wt})
Intermediate	<ul style="list-style-type: none"> Other cytogenetic and molecular abnormalities‡ (<i>FLT3</i>-ITD^{pos} and/or <i>NRAS</i>^{mut} and/or <i>KRAS</i>^{mut}; <i>TP53</i>^{wt})
Adverse	<ul style="list-style-type: none"> Mutated <i>TP53</i>

This classification does not apply to patients who have received prior treatment with an HMA or who have progressed from MPN.

† consider germline testing

*if treated with Aza/Ivosidenib

‡For many cytogenetic and molecular abnormalities, single or as coaberrations, no data are currently available; they are tentatively categorized as favorable and intermediate-risk depending on the absence or presence of activating signaling gene mutations.



ELN 2024 Less Intensive

Favorable-risk group	Median OS, months	Reference
Mutated <i>NPM1</i> (<i>FLT3-ITD</i> ^{neg} , <i>NRAS</i> ^{wt} , <i>KRAS</i> ^{wt} , <i>TP53</i> ^{wt})	39	4
Mutated <i>IDH2</i> (<i>FLT3-ITD</i> ^{neg} , <i>NRAS</i> ^{wt} , <i>KRAS</i> ^{wt} , <i>TP53</i> ^{wt})	37	4
Mutated <i>IDH1*</i> (<i>TP53</i> ^{wt})	29	6,17
Mutated <i>DDX41</i>	>24	3,13
AML with MR gene mutations (<i>FLT3-ITD</i> ^{neg} , <i>NRAS</i> ^{wt} , <i>KRAS</i> ^{wt} , <i>TP53</i> ^{wt})	23	4
Intermediate-risk group		
AML with MR gene mutations (<i>FLT3-ITD</i> ^{pos} and/or <i>NRAS</i> ^{mut} and/or <i>KRAS</i> ^{mut} ; <i>TP53</i> ^{wt})	13	4
Other cytogenetic and molecular abnormalities (<i>FLT3-ITD</i> ^{pos} and/or <i>NRAS</i> ^{mut} and/or <i>KRAS</i> ^{mut} ; <i>TP53</i> ^{wt})	12	4
Adverse-risk group		
Mutated <i>TP53</i>	5-8	3,4,7,10,14-16

4 Döhner, et al. Genetic risk stratification and outcomes among treatment-naïve patients with AML treated with venetoclax and azacitidine. *Blood*. 2024

6 Montesinos, et al. Ivosidenib and azacitidine in IDH1-mutated acute myeloid leukemia . *N Engl J Med*. 2022

3 Jahn, et al. Clinical impact of the genomic landscape and leukemogenic trajectories in non-intensively treated elderly acute myeloid leukemia patients. *Leukemia*. 2023



Evolving Classifications and more to come

FAB-Classification 1976

FAB-Classification update 1989

WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues 1997

WHO Classification 2001

WHO Classification 2008

Revised 4th WHO Classification 2017

5th WHO Classification 2022

NCCN v1.2025

ELN 2010

ELN 2017

ELN 2022

ELN 2024 less intensive

ICC 2022



Choosing the right MRD assay

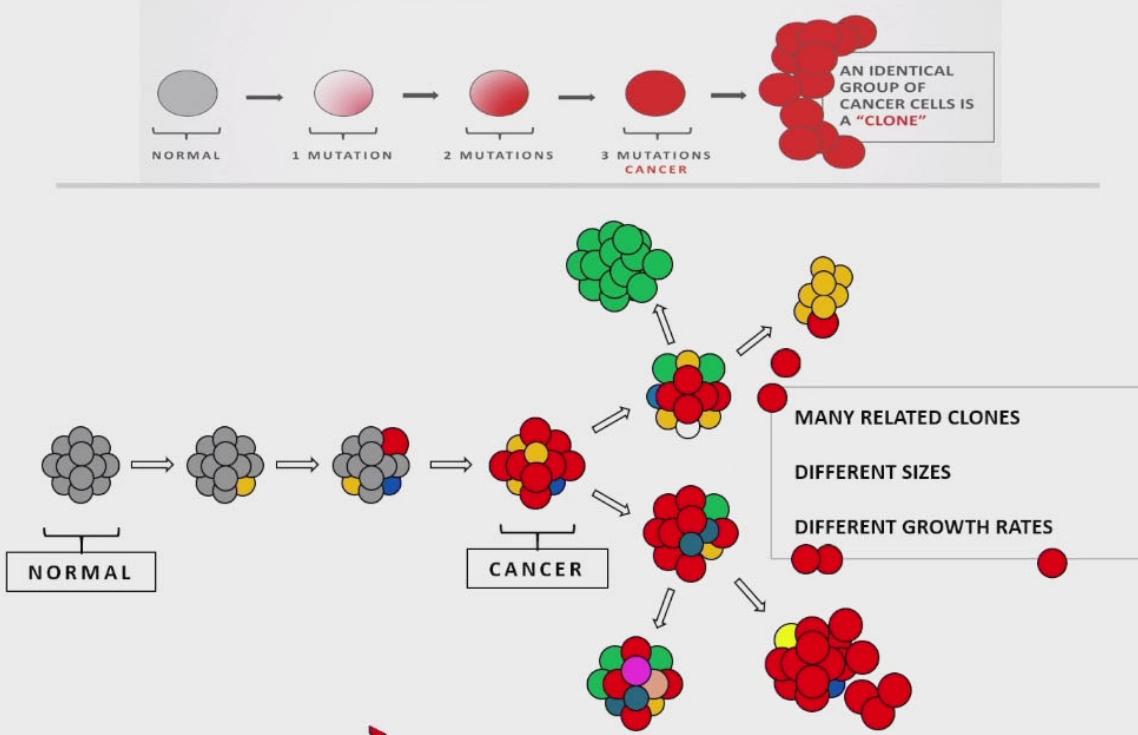
ELN risk group	Genetic subgroup	MRD assay in non-transplanted patients	MRD assay after alloHCT
Favorable	NPM1 mut RUNX1/RUNXT1 or CBFB/MYH11 CEBPA bZIP inframe	qPCR qPCR MFC #1570	qPCR qPCR Not established
Intermediate	FLT3-ITD NPM1wt FLT3-ITD NPM1mut MLLT3::KMT2A Other	FLT3-NGS or MFC qPCR + FLT3-NGS MFC or qPCR MFC	FLT3-NGS or MFC qPCR (FLT3-NGS) qPCR (MFC, NGS) MFC (or NGS)
Adverse	Fusion genes -5 or del(5q); -7; -17/abn(17p) Complex karyotype Myelodysplasia-related gene mutations TP53	qPCR or MFC MFC MFC MFC	qPCR or MFC (or NGS) MFC (or NGS) MFC (or NGS) MFC (or NGS)



American Society of Hematology

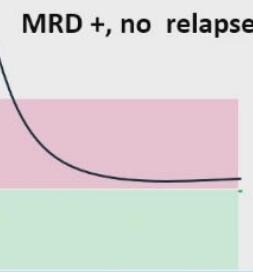
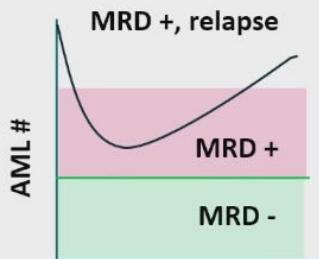
Heuser M&M 2024

Cancer often has many clones-making of the cancer ecosystem



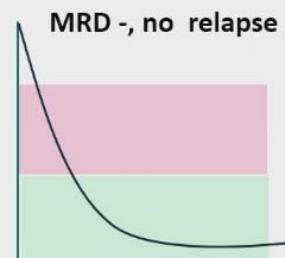
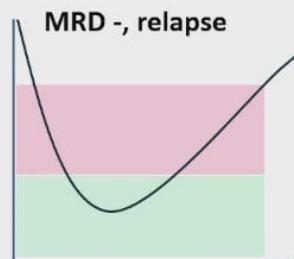
Four possible states of MRD and relapse

- This makes sense
- Persistent clone?
- New clone?



- Some clones more "tolerable?"
- Persistence of "CHIP" mutations?
- Mutation in lymphoid lineage?
- GVL?

- Need a better test?
- Clonal evolution and loss of markers?
- New clone?

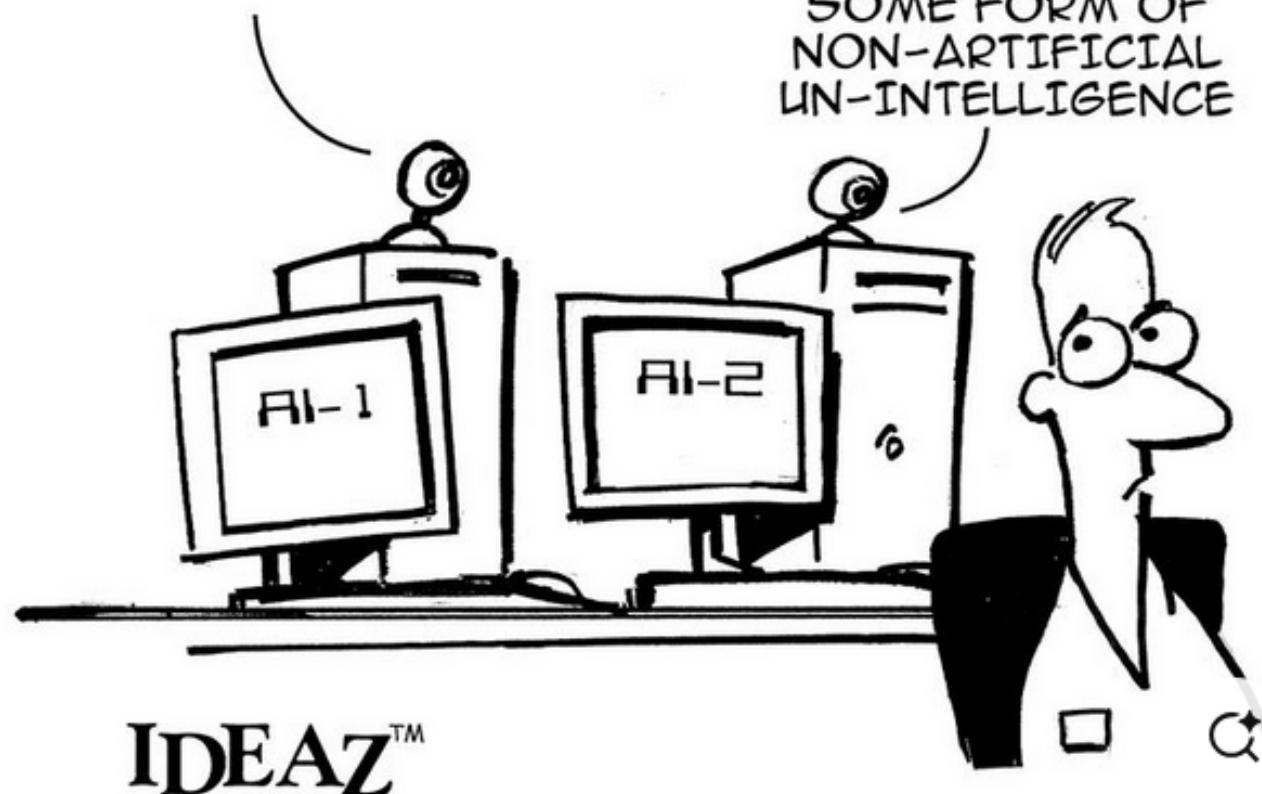


- This makes sense



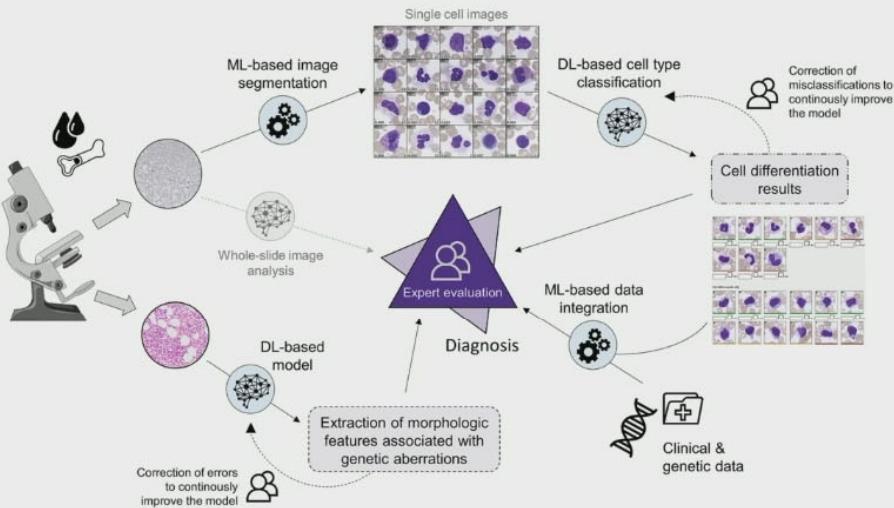
WHAT IS THAT THING
SEATED NEXT TO US?

I'M NOT SURE - BUT
IT APPEARS TO BE
SOME FORM OF
NON-ARTIFICIAL
UN-INTELLIGENCE



IDEAZ™

Applications in Cytomorphology



Important questions:

What is the intended use? Augment or replace humans?

High relevance for required accuracy of output

Walter W. et al. Blood Reviews 2022

Guilherme de Almeida J. et al., Nature Communications 2023

Reproduce current diagnostic approach
cell (pre)classification reviewed by an expert to optimize the diagnostic workflow

Identify new features for disease classification
e.g. to distinguish SF3B1-mutant MDS from other MDS using cytomorphology and blood counts alone

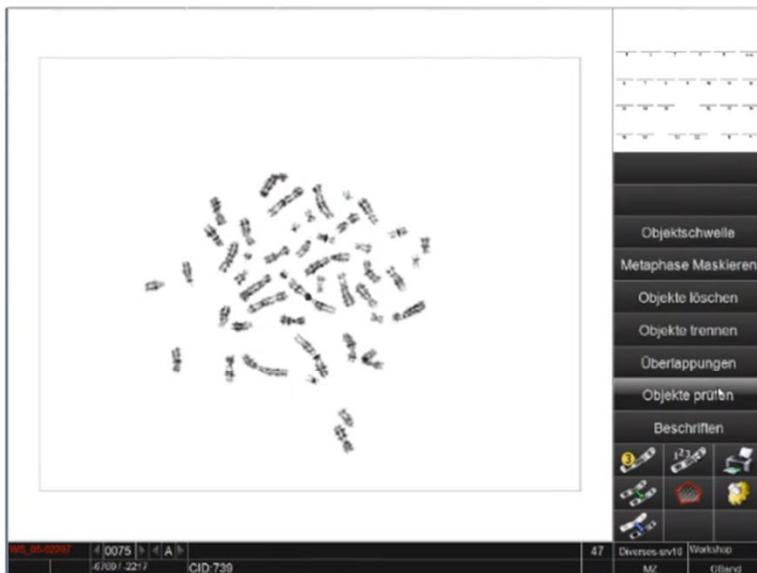


Automated karyotyping

Normal karyotype



Experienced technician



三

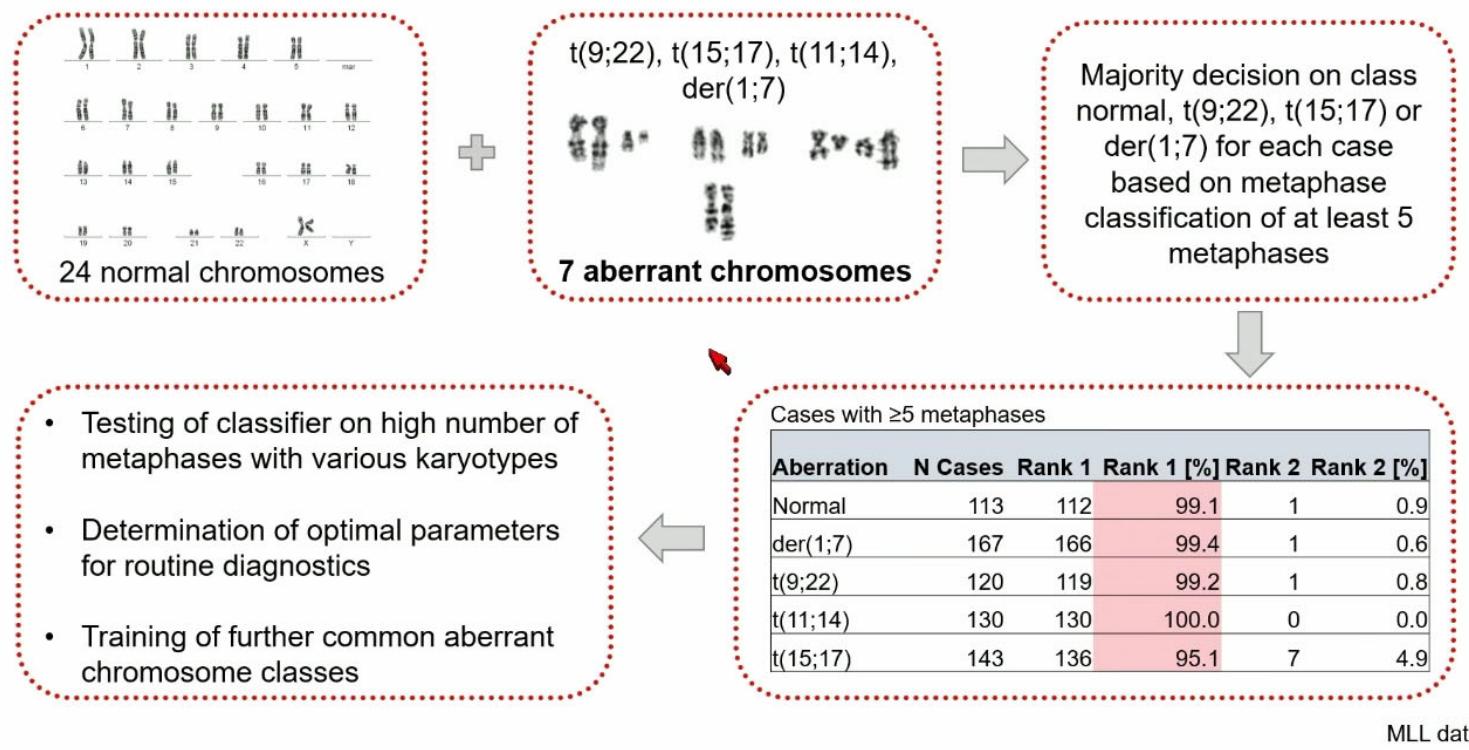
A man in a dark suit and glasses is speaking at a podium. He is wearing a light-colored shirt and a patterned tie. The podium has a red circular logo with a white stylized 'S' or 'A' in the center. The text "65th ASH® Annual Meeting" is printed below the logo. In the background, there are red curtains.

65th ASH® Annual Meeting

MLL data



Next steps – Identification of chromosome aberrations

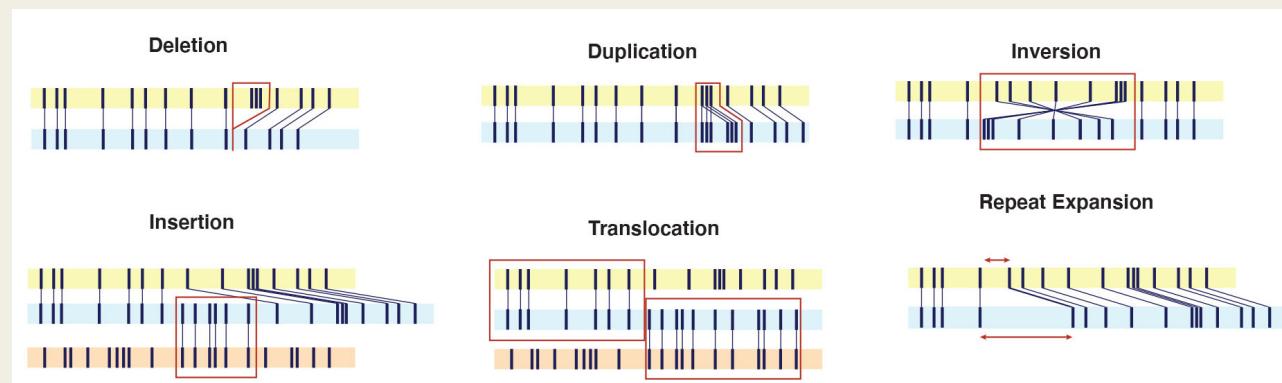
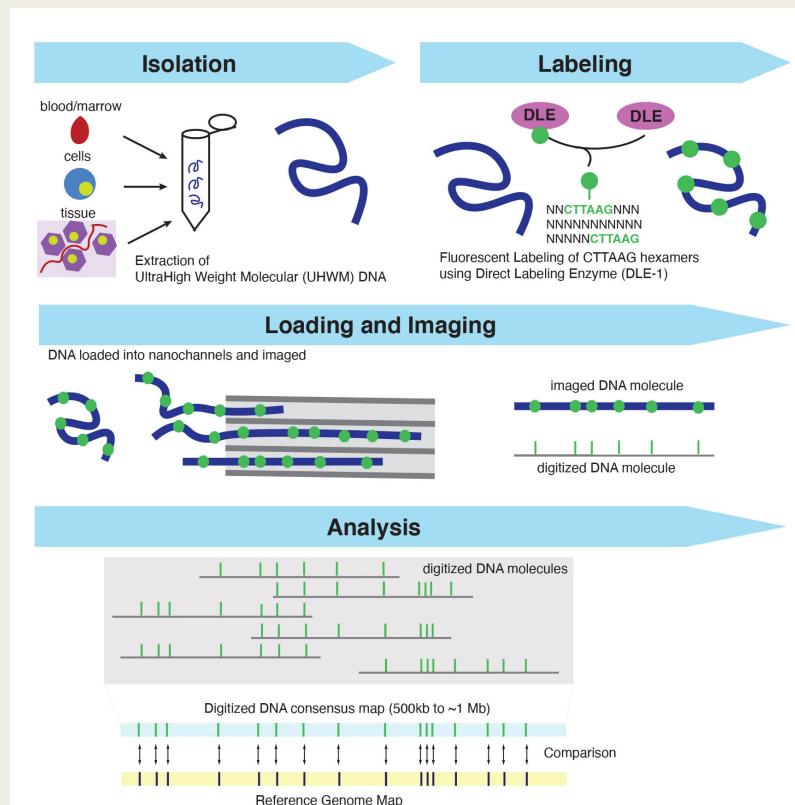


New Method Edging Towards Routine Diagnostics ?

– Optical Genome Mapping



Optical Genome Mapping -



OPTICAL GENOME MAPPING (OGM) AS STANDARD OF CARE FOR ACUTE LEUKEMIA DIAGNOSIS: DIAGNOSTIC AND CLINICAL IMPACTS 10 MONTHS POST-IMPLEMENTATION



E. McGinnis MD FRCPC^{1,2}, R. Stubbins, MD MSc FRCPC³, D. Li PhD FCCMG^{1,2}, Z. Hamadeh PhD^{1,2}, and T. Spence PhD FCCMG DABMG^{1,2}

1. Department of Pathology and Laboratory Medicine, Vancouver General Hospital, Vancouver, BC, Canada
2. Department of Pathology and Laboratory Medicine, University of British Columbia, Vancouver, BC, Canada
3. Leukemia/Bone Marrow Transplant Program of British Columbia, British Columbia Cancer Agency, Vancouver, Canada



THE UNIVERSITY
OF BRITISH COLUMBIA



Poster Session Online
Presented by Eric McGinnis

Download
OGM was
available
on mobile
device



Amgen Society for Bone Marrow
Transplantation and Cellular Therapy
Annual Meeting
May 18-21, 2023, Boston, MA, USA

BACKGROUND

- Optical genome mapping (OGM) is a novel method for detecting chromosomal structural and copy number variants by imaging and mapping ultra-long fluorescently-labelled DNA.
- OGM detects cancer-associated variants with hundreds- to thousands-fold higher resolution than standard cytogenetic methods (karyotyping/CG/FISH).
- Our clinical laboratory in British Columbia, Canada's largest tertiary care hospital implemented OGM as front-line standard-of-care testing for all newly diagnosed acute leukemias in parallel with CG/FISH.

AIM

- To prospectively describe performance and impact on diagnosis of OGM in acute leukemia diagnostic genomic profiling.

METHODS

- OGM was performed per manufacturer protocols on bone marrow aspirate or, when unavailable, blood using a Bionano Genomics instrument and Rare Variant Analysis informatics (paired with De Novo or Guided Assembly for ALL).
- Detected variants were filtered and reported using population thresholds and a custom 277-gene/region file and validated laboratory protocols for variant identification and classification in indication-specific contexts.
- All patients received concurrent CG/FISH (limited) analysis as routinely indicated.

RESULTS

- 90 adults (mean age 55 years, 47% female) with acute leukemia had OGM performed at diagnosis, including 62 with acute myeloid leukemia (AML), 27 with lymphoblastic leukemia, including 22 B lineage (B-ALL) and 5 T lineage (T-ALL), and one mixed phenotype acute leukemia (T/myeloid).
- OGM provided adequate data for reporting for 86 individuals (98%) with a mean time to result availability of 11.8 calendar days following sample procurement.
- Both instances of OGM failure resulted from inadequate bone marrow aspirate volume and insufficient circulating disease for processing.

Table 1. Variants identified by OGM and impact on pathologic diagnosis and risk stratification, among cases for which any detectable OGM or cytogenetic abnormalities were identified

	AML	B-ALL	T-ALL
OGM variants, mean	1.4	4.7	3
Additional variants versus CG/FISH, mean	0.6	3.4	3
Change in pathologic diagnosis, frequency	9 (15%)	1 (5%)	0
Change in risk, frequency	12 (19%)	11 (50%)	0

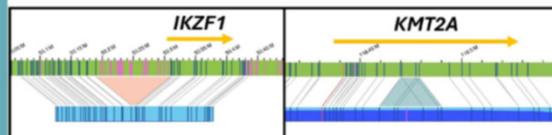


Figure 1. Select cryptic risk-modifying abnormalities identified solely by OGM. Additional variants predicting increased risk were identified in 12 individuals with AML and 11 individuals with B-ALL; these abnormalities included *IKZF1* deletion (left), *KMT2A* partial tandem duplication (right), complex karyotype (not shown), and cryptic deletion of *RUNX1* (not shown)

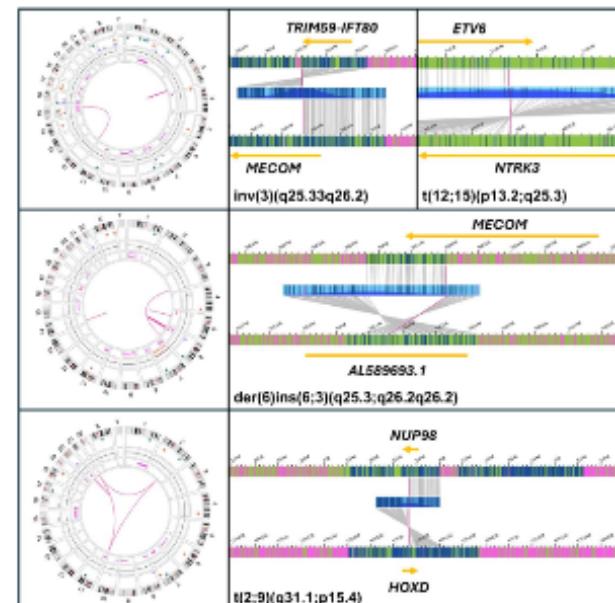


Figure 2. Select cryptic novel driver rearrangements identified by OGM. OGM changed 11% of pathologic diagnoses: 9 AML (3 MECOM and 2 NUP98 rearrangements, 2 *RUNX1* deletions, an unbalanced Tq rearrangement, and a germline CHEK2 deletion) and 1 B-ALL (ZNF384 rearrangement). 4 novel rearrangements were defined, including MECOM::IL12A-AS1/TRIM59-IFT80 resulting from inv(3) in AML (top), MECOM::AL589693.1 resulting from unbalanced ins(6;3)(q25.3;q26.2q26.2) in oligoblastic AML (middle), and NUP98::HOXD resulting from t(2;9) in AML (bottom)

CONCLUSION

OGM expands the spectrum of detectable cytogenomic abnormalities in acute leukemias, resulting in meaningful disease reclassification and enhanced risk stratification in these diseases, and can permit patients access to previously inaccessible targeted therapies. Data regarding the prognostic impact and actionability of recurrent variants newly detectable or characterizable by OGM are urgently needed to maximally leverage this transformative technology.

Contact: eric.mcginnis@vch.ca

SAT-1150
ERIC MC GINNIS

und am Ende....

Vielen Dank für
Ihre Aufmerksamkeit